Latent Block Model for Overdispersed Count Data Application in Microbial Ecology

J. Aubert Joint work with S. Robin, S. Schbath

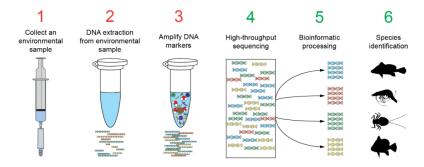


MathsForGenomics Evry

Metabarcoding

A way of studying diversity data for entire communities from environmental samples.

Operational Taxonomic Unit (OTU) are identified by sequencing a standardized region of DNA.



A typical metagenomic experiment

Amplicon-based sampling. Consider

- ▶ *n* different (bacterial, fungal, ...) species / OTU and
- m different samples / patients / media / conditions.

NGS provides

- Y_{ij} = number of reads from species *i* in sample *j*
 - \propto abundance of species *i* in sample *j*

Question. Can we exhibit some patterns in the distribution of the species abundances across samples?

Bi-clustering problem

Rephrased problem : Find

- groups of species having similar abundance profile across the samples and
- groups of samples histing the different species in similar proportions.

Bi-clustering problem

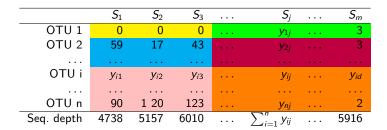
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- groups of samples histing the different species in similar proportions.

Bi-clustering problem : Simultaneously determine

- row clusters and
- column clusters
- in a $n \times m$ matrix of counts.

Bi-clustering problem



 y_{ij} = number of sequences from sample *j* assigned to Operational Taxonomic Unit (OTU) *i*.

Approach

Model-based clustering :

```
\rightarrow \mathsf{LBM} = \mathsf{Latent} \; \mathsf{Block}\text{-}\mathsf{Model}
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(Govaert and Nadif, 2005; Brault and Mariadassou, 2015)

Approach

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Specificities of NGS data :

- count data,
- over dispersed (wrt Poisson),
- ▶ with heterogeneous sampling effort (= sequencing depth),
- with high variation among the species abundances,
- possibly with replicates.

Latent Block Model

Bi-clustering. K species groups, G sample groups

- ▶ Z_i = group to which species *i* belongs to ($\in \{1, ..., K\}$);
- W_j = group to which sample j belongs to ($\in \{1, ..., G\}$)

both latent = hidden = unobserved.

 \rightarrow Incomplete data model

Latent Block Model

Bi-clustering. K species groups, G sample groups
Z_i = group to which species i belongs to (∈ {1,...K});
W_j = group to which sample j belongs to (∈ {1,...G})
both latent = hidden = unobserved.

 \rightarrow Incomplete data model

Ex : Poisson LBM.

 $\begin{array}{cccc} (Z_i) \mbox{ iid } \sim & \pi & (\mbox{species prop.}) \\ (W_j) \mbox{ iid } \sim & \rho & (\mbox{sample prop.}) \\ (Y_{ij}) \mbox{ indep } |(Z_i); (W_j) & \sim & \mathcal{P}(\lambda_{Z_iW_j}) \end{array}$

Does not accommodate for NGS data specificities.

Over-dispersion

Negative-binomial. Most popular distribution of NGS counts :

$$Y \sim \mathcal{NB}(\lambda, \phi)$$
 $\mathbb{E}(Y) = \lambda,$ $\mathbb{V}(Y) = \lambda(1 + \phi\lambda) \geq \lambda.$

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Gamma-Poisson representation. Take $a = 1/\phi$ and draw

 $U \sim \mathcal{G}am(a, a), \quad Y \mid U \sim \mathcal{P}(\lambda U) \quad \Rightarrow \quad Y \sim \mathcal{NB}(\lambda, \phi).$

Negative binomial = Poisson with latent Gamma

 \rightarrow Incomplete data model (Y is observed, U is not).

LBM for metagenomic data

Hidden layer :

LBM for metagenomic data

Hidden layer :

Observed counts : (interest of model-based approaches)

$$Y_{ij} \mid Z, W, U \sim \mathcal{P}\left(\mu_i \ \nu_j \ \alpha_{Z_i W_j} \ U_{ij}\right)$$

where

- $\blacktriangleright \mu_i$: mean abundance of species *i*
- ν_j : sequencing depth in sample *j* (fixed)

 $\triangleright \alpha_{kg}$: interaction term between group species k and sample group g.

LBM for metagenomic data

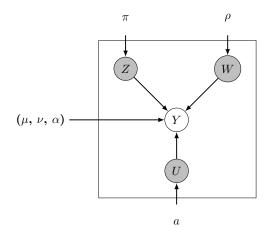


FIGURE – The proposed over-dispersed Poisson LBM presented as a directed graphical model. Legend : observed variables (filled white), latent variables (filled gray), parameters are outside the box.

Inference

Aim : Retrieve

- ▶ Z_i = species group, or at least $P(i \in k|Y)$;
- W_j = sample group, or at least $P(j \in g|Y)$;

and estimate the interaction parameter $\alpha = (\alpha_{kg})$.

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Which means (maximum-likelihood approach)

- Compute p(Z, W, U|Y);
- Maximize log $p_{\theta}(Y)$, where $\theta = (\alpha, \mu)$.

Most popular algorithm : EM (Dempster et al., 1977).

Variational approximation

Species group Z_i and sample group W_j are not independent given Y_{ij} $\rightarrow p(Z, W, U \mid Y)$ intractable

Variational approximation (Jordan, 1999). Find

 $\begin{array}{lll} & \widetilde{p}(Z,W,U) &\simeq & p(Z,W,U|Y) \\ \text{such that} & & \widetilde{p}(Z,W,U) &= & \widetilde{p}_1(Z) \ \widetilde{p}_2(W) \ \widetilde{p}_3(U) \end{array}$

(mean-field approximation).

ightarrow Variational EM (VEM) algorithm provide a lower bound

$$J(Y, \widetilde{p}, \widehat{ heta}) \leq \log p_{\widehat{ heta}}(Y).$$

Penalized 'likelihood' criteria

Penalized criterion. log $p_{\hat{\theta}}(Y)$ intractable

$$\log p_{\widehat{\theta}}(Y) - \operatorname{pen}(p_{\widehat{\theta}}) \longrightarrow J(Y,\widetilde{p},\widehat{\theta}) - \operatorname{pen}(p_{\widehat{\theta}})$$

BIC & ICL. $\mathcal{H} = \text{entropy}$ $\text{pen}_{BIC} = [(K-1)\log n - (G-1)\log m - KG\log(nm)]/2$ $\text{pen}_{ICL_1} = \text{pen}_{BIC} + \mathcal{H}(\widetilde{p}_Z) + \mathcal{H}(\widetilde{p}_W)$ (classification entropy)

Model comparison

Likelihood ratio for nested models.

 $\mathcal{M} \subset \mathcal{M}',$ the likelihood ratio is defined as

$$LR(\mathcal{M}, \mathcal{M}') = 2\left[\log p(\mathbf{Y}; \widehat{\theta}_{\mathcal{M}'}) - \log p(\mathbf{Y}; \widehat{\theta}_{\mathcal{M}})\right].$$

 $\begin{array}{l} \mbox{Interest of block structure.} \\ \mathcal{M}_{\min} := \mathcal{M}_{1,1} \subset \mathcal{M}_{K,G} \subset \mathcal{M}_{\max} := \mathcal{M}_{n,m} \end{array}$

Lower bounds for likelihood ratios.

(a):
$$LR(\mathcal{M}_{\min}, \mathcal{M}_{K,G}) \geq 2\left[\mathcal{J}(\mathbf{Y}, \widehat{q}_{K,G}, \widehat{\theta}_{K,G}) - \log p(\mathbf{Y}; \widehat{\theta}_{1,1})\right],$$

$$(b): \quad LR(\mathcal{M}_{K,G},\mathcal{M}_{\max}) \leq 2\left[\log p(\mathbf{Y};\widehat{\theta}_{n,p}) - \mathcal{J}(\mathbf{Y},\widehat{q}_{K,G},\widehat{\theta}_{K,G})\right].$$

Three 16S or 18S rRNA amplicon-based datasets

- MetaRhizo : plants and bacteria communities living in their rhizosphere (collab. C. Mougel, INRA Rennes)
- Oak powdery mildew : bacteria and fungi including *Erysiphe* alphitoides living in the phyllosphere (collab. C. Vacher, INRA Bordeaux)
- Macaroni : microbial community assembly in soil (collab. L. Philippot, A. Spor, INRA Dijon)
- Aim : to understand the structure of these relationships

Meta-rhizo

Dataset : Medicago truncatula rhizosphere.

▶ *n* = 288 bacteria (genus)

• m = 483 samples = rhizosphere of different plants (genotypes)

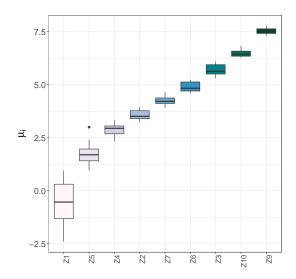
The total counts per sample go from 29410 to 33840 number of sequences.

19.2% of data are null

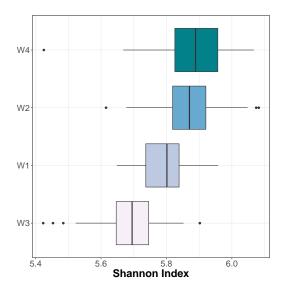
Range from 0 to 5084 with a median = 9 and mean = 110

Results :

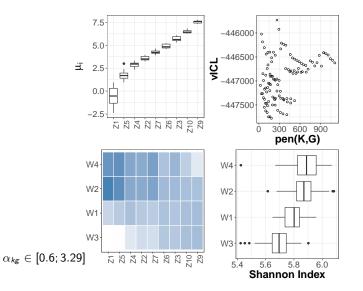
- $\widehat{K} = 10$ groups of bacteria
- $\hat{G} = 4$ groups of samples



Despite ν_j , bacteria groups correspond to abundance groups.



Plant groups corresponds to diversity levels (Shannon index).



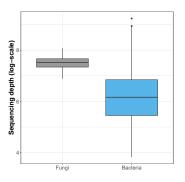
Goodness of fit.

 T_{ABLE} – MetaRhizo data. Goodness-of-fit. LR is the likelihood ratio statistic as defined in Section and df stands for difference in terms of free parameters.

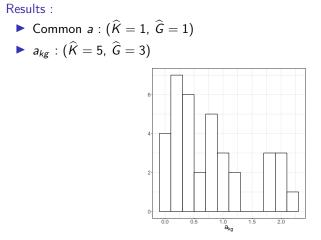
\mathcal{M},\mathcal{M}'	$LR(\mathcal{M},\mathcal{M}')$	df	$LR(\mathcal{M},\mathcal{M}')/df$
$\mathcal{M}_{min}, \mathcal{M}_{\mathit{KG}}$	37804.75	40	945.12
$\mathcal{M}_{\mathit{KG}}, \mathcal{M}_{max}$	143881	139064	1.03

Dataset : Pathobiome of the Erysiphe alphitoides (Jakuschkin et al. 2016).

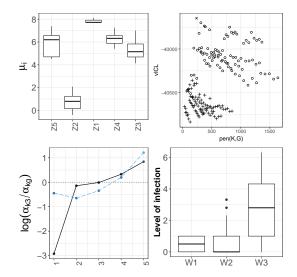
- ▶ n = 114 = E. alphitoides +47 fungal +66 bacterial otus
- m = 116 leaves from 3 trees (resistant, intermediate, susceptible)
- 34% of data are null
- Range from 0 to 2228 (median = 2; mean = 24.17)



$$ightarrow 2
u_j = \left(
u_j^{\text{bact}},
u_j^{\text{fung}}
ight)$$



 $\alpha_{kg} \in [0.22; 2.14]$ (ratio from 1 to 9.6).



Comments :

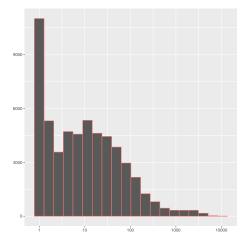
- Heterogeneous over-dispersion parameters (a_{kg}) ,
- ► Groups reveal the abundance of *E. alphitoides* (pathogene)

MicrobiAl Community Assembly Rules and functiONIng

Aim : Identify biotic interactions between microbial groups using a targeted subtractive approach by removal and enrichment of specific microbial groups

Data : After filtering steps, 353 OTUs and 347 biological samples (10 treatments)

▶ 54% of data are null, Mean = 35.3, Max = 10598.

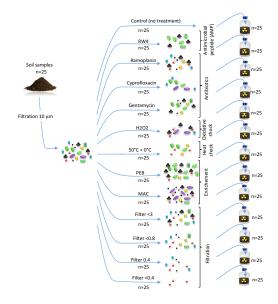


J. Aubert

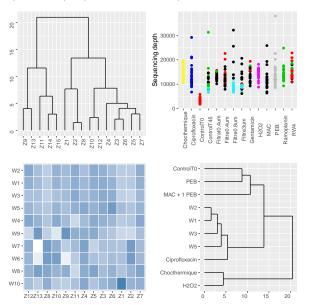
Approach and methods

- 1. Ten time dilution of soil suspension filtered at 10 $\mu{\rm m}$ to focus on dominant bacterial groups.
- 2. Removal, killing or preventing the growth of specific groups
 - according to their cell size using filtration (4 size classes)
 - by incubating the soil suspension with (i) antibiotics targeting different groups and (ii) group specific antimicrobial peptides
 - according to the membrane properties by subjecting the soil suspension to osmotic and heat shocks
 - enrichment by incubating the soil suspension with inhibitors
- 3. For each treatment : inoculation into 25 microcosms containing sterilized soils.
- 4. Collect after 45 days for molecular and activity analyses.
- 5. Illumina Miseq sequencing
- 6. Bioinformatic annalysis with house pipeline (A. Spor)

Experimental Design



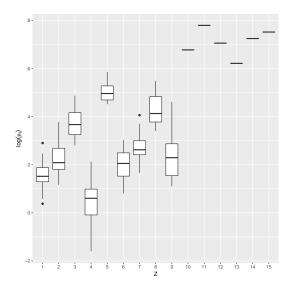
Selected latent block model Common a ($\hat{a} = 0.32$) : ($\hat{K} = 15$, $\hat{G} = 10$)



Description of groups in columns

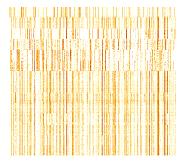
	W_1	W_2	W_3	W_4	W_5	W_6	W_7	W_8	W_9	<i>W</i> ₁₀
Chocthermique	0	0	0	0	0	0	25	0	0	0
Ciprofloxacin	1	0	0	24	0	0	0	0	0	0
ControlT0	0	0	0	0	0	0	0	0	0	25
ControlT45	6	0	16	0	3	0	0	0	0	0
Filtrat0.4um	13	10	2	0	0	0	0	0	0	0
Filtre0.4um	17	8	0	0	0	0	0	0	0	0
Filtre0.8um	5	0	0	0	20	0	0	0	0	0
Filtre3um	19	1	0	0	5	0	0	0	0	0
Gentamicin	17	4	3	0	0	0	0	0	0	0
H2O2	1	0	0	0	0	23	0	0	0	0
MAC	0	0	0	0	0	0	0	0	25	0
PEB	0	0	0	0	0	0	0	24	1	0
Ramoplanin	0	0	24	0	0	0	0	0	0	0
RW4	4	18	3	0	0	0	0	0	0	0

Groups of bacteria

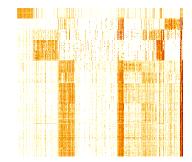


Heatmap

Before

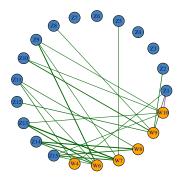


After



Network representation

- One vertex = one group of microorganism (Z_i in blue) or one group of soil (W_i in orange)
- lncidence matrix : use of α_{kg} matrix (abs. value > 1)
- Edge color : green for negative, purple for positive interactions



Discussions

Summary

- Parsimonious and complex model enables us to reduce data dimension
- ICL criteria to select number of groups
- Parameters biologically interpretable
- cobiclust R package

Possible extensions

Comments

- Dispersion parameter
- Normalization
- Zero-inflation

Acknowledgments

For experiments, datasets and biological expertise







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For the statistical part

S. Robin S. Schbath S. Ouadah

References

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